# Nutritional rehabilitation of malnourished rats by di- and tripeptides: nitrogen metabolism and intestinal response

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An enzymic hydrolysate (DTP) rich in di- and tripeptides (75%) was prepared from bovine plasma proteins (BP). Protein-calorie malnourished rats were subjected to refeeding tests with diets having the same nitrogen and caloric contents and differing only in the molecular form of the nitrogen supply: BP, DTP, or an equivalent amino acid mixture (AAM). Growth, food intake, and food efficiency were measured after an eight-day period of rehabilitation; nitrogen balance and digestibility were also determined. Animals refed the DTP diet exhibited the best nutritional parameters, AAM failed to support any rehabilitation. In the intestine measurements of mucosal enzyme activities and morphometric studies strongly suggested the beneficial effect of DTP as indicated by increased both aminopeptidase N level and villus height. The results showed an adaptive response of rats to the molecular form of the nitrogen supply and demonstrated the superior efficacy of DTP for a rapid recovery from protein-calorie malnutrition.

Keywords: refeeding; di and tripeptides; intestinal enzymes and morphology; nutritional adaptation

# Introduction

Partial protein hydrolysates or equivalent amino acid mixtures are widely used for medical and therapeutic uses, either for enteral or parenteral nutrition<sup>1,2</sup> in the rehabilitation of undernourished patients with or without intestinal diseases. Thus, it seems preferable to test the efficacy of such products using similar animal models, ie, protein-calorie malnourished rats (low protein level and reduced food intake), rather than starved animals (absence of luminal nutrients).

Undernutrition has been shown to cause lowered

rapid restoration of intestinal physiology. 17-20 However, many of these numerous studies have been performed using either acute starvation, protein-free diets, or a litter expansion technique. Moreover, no attempt has been made to investigate the effects of diets differing only in the molecular form of the nitrogen source supplied on the nutritional rehabilitation of undernourished rats. The aim of this work is to determine if rehabilitation is more effective using diets with hydrolyzed protein or amino acids, compared with using diets containing whole protein; thus, we have chosen to conduct this study to examine the effects of bovine plasma proteins, a partial enzymic hydrolysate of these proteins, and a free amino acid mixture equivalent to the hydrolysate, on the intestinal response (ie, enzyme activity and morphology) of

small intestinal function<sup>3-6</sup> associated with decreased mucosal enzyme activities<sup>7-9</sup> and severely impaired

morphology. 10-16 Resuming food intake results in a

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malnourished animals. In addition, our attention was

turned on their overall nutritional adaptation.

### Materials and methods

# Partial hydrolysate (DTP) of plasma proteins (BP)

Blood was collected in a slaughter house. Spray-dried bovine plasma proteins (BP) were hydrolysed by an alkaline bacterial serine proteinase according to Bressollier et al. <sup>21</sup> Resulting peptides were continuously extracted by a membrane ultrafiltration process (molecular weight cut off: 1,000 daltons). Then, each peptide class, characterized by its size, was quantified on the basis of its amino acid content after separation by ligand exchange chromatography using Cu(ll)-modifield silica gel. The relative distribution, as moles percentage, was: amino acids, <5; dipeptides,  $40 \pm 4$ ; tripeptides,  $35 \pm 4$ ; larger peptides (chain length <6 residues),  $20 \pm 2$  (Verneuil et al., unpublished data). This hydrolysate, rich in di- and tripeptides, was named DTP. The amino acid composition of BP and DTP is further described and reported in *Table 1*.

### Animal model and diets

Twenty male rats (Sprague-Dawley strain) weighing 106 ± 3 g were housed in stainless steel metabolic cages with urine-feces separators in an animal room kept at 25° C with a 12-hr light: dark cycle. They were fed a diet containing casein (2.0 g of protein per 100 g of diet) and their food intake was limited to an average of 5.5 g/day in order to achieve body weight loss. Water was given ad libitum. On day 16 five animals were killed (controls). The remaining rats were then divided in three equal groups (five rats per group) based on average body weight; they were switched to isoenergetic and isonitrogenous diets (2.4%) N) containing either BP, DTP, or an amino acid mixture (AAM) as the sole nitrogen source (2.4% N  $\times$  6.25, ie, 15% protein level). Refeeding lasted for 8 days. Rats were killed on the day 9. Although the composition of the diets was expressed as percentage on a dry basis (Table 2), they were given to rats on a gruel form containing 60% of moisture, to avoid food spillage. Proteins, peptides, and amino acids (Merck-Clevenot, Nogentsur-Marne, France) were in powder form and mixed with the rest of the diet. In diets containing AAM, all the nitrogen was supplied as a mixture of cristalline L-amino acids corresponding to the composition of the DTP mixture. Every day food con-

 Table 1
 Amino acid composition of the nitrogen sources

	Composition (amino acid %)	
Aminoacid	BP	DTP
Aspartic acid	7.21	9.70
Threonine	6.73	6.98
Serine	7.03	7.53
Glutamic acid	14.13	14.66
Proline	4.09	4.04
Glycine	3.45	3.38
Alanine	4.85	5.12
Cysteine	1.37	1.44
Valine	6.73	6.80
Methionine	1.01	1.04
Isoleucine	3.10	3.01
Leucine	9.05	8.85
Tyrosine	4.80	4.79
Phenylalanine	4.88	4.67
Tryptophane	1.79	1.35
Lysine	9.05	8.73
Histidine	2.44	2.46
Arginine	5.30	5.36

**Table 2** Composition of the experimental diets expressed as g/100g of diet (dry basis)

		Dietary groups		
	Malnutrition		Refeeding	
Ingredients	Control	BP	DTP	AAM
Casein (13.8% N)	2.32			
DL-methionine <sup>a</sup>	0.04	_	_	
BP (14.1% N)	_	17.05		
DTP (15.5% N)	_	*****	15.52	
AAM (16.0% N)			_	15.00
Starch	68.37	58.06	59.13	59.50
Glucose	4.88	4.15	4.22	4.25
Cellulose	4.88	4.15	4.22	4.25
Fat	7.81	6.64	6.76	6.80
Salt mixture <sup>b</sup>	10.75	9.12	9.30	9.35
Vitamin mixture <sup>c</sup>	0.98	0.83	0.84	0.85
Protein <sup>d</sup> and energy level in each diet				
$N \times 6.25$	2.0	15.0	15.0	15.0
Kcal (calcd) <sup>e</sup>	392	393	393	393
KJ (calcd)	1638	1643	1643	1643

<sup>&</sup>lt;sup>a</sup> Casein was supplemented with 2% DL-methionine.

sumption and body weight were individually recorded. During the last 3 days of the malnutrition and refeeding periods the urine and feces were collected for nitrogen balance and digestibility studies.

### Experimental procedure

At the end of each experimental period rats were killed by cervical dislocation and the abdomens rapidly opened. The subsequent steps were carried out at 4° C. The intestine was removed intact from the pylorus to the ileocecal valve and three standard segments were excised: duodenum (from pyloric valve to Treitz's ligament), middle jejunum (consisting of the portion 30-45 cm distal to the Treitz's ligament) and ileum (consisting of the portion 15-30 cm proximal to the ileocecal valve). After flushing with ice-cold saline, the intestinal mucosae were scraped and homogenized in the same medium, then stored at -80° C until assays. Brush border alkaline phosphatase (AP, duodenum) and aminopeptidase N (AN, ileum) activities were assayed according to Bessey et al. 22 and Maroux et al., 23 respectively. Protein was determined by the Lowry procedure. 24

# Morphometric study

Immediately after the sacrifice, I cm specimens were taken from identical parts of the digestive tract (beginning of the middle jejunum), fixed in Bouin's solution, embedded in paraffin, cut perpendicular to the longitudinal axis and stained with

<sup>&</sup>lt;sup>b</sup> Salt mixture consisted of (g/kg): calcium dihydrogen phosphate: 430.0; potassium chloride: 100.0; sodium chloride: 100.0; magnesium chloride: 50.0; magnesium sulfate: 50.0; ferric oxide: 30.0; manganese sulfate: 2.45; zinc sulfate: 2.00; cupric sulfate: 0.50; cobalt sulfate: 4.10<sup>-3</sup>; potassium iodide: 8.10<sup>-3</sup>.

<sup>&</sup>lt;sup>c</sup> Composition expressed in international units or g per kg of vitamin mix: retinyl acetate: 1980,000 IU; cholecalciferol: 600,000 IU; DL-tocopheryl acetate: 17.0; menadione: 4.0; thiamin-HCl: 2.0; riboflavin: 1.5; calcium pantothenate: 7.0; pyridoxin-HCl: 1.0; inositol: 15.0; cyanocobalamine: 5.10<sup>-3</sup>; ascorbic acid: 80.0; nicotinic acid: 10.0; choline-HCl: 136.0; folic acid: 0.5; p-aminobenzoic acid: 5.0; D-biotin: 3.10<sup>-2</sup>.

 $<sup>^{\</sup>rm d}$  Calculated as: grams of nitrogenous ingredient  $\times$  % nitrogen  $\times 6.25.$ 

 $<sup>^{\</sup>rm e}$  Calculated as: total carbohydrates  $\times$  4 kcal/g; fat  $\times$  9 kcal/g; protein  $\times$  4 kcal/g.

Table 3 Effects of malnutrition on nutritional parameters<sup>a</sup>

Parameters	Controls
Initial body weight (g) Final body weight (g) Protein intake (g/day) Caloric intake (kcal/day)	102.60 ± 1.90 73.40 ± 3.20 0.11 ± 0.01 21.95 ± 0.51
N intake (mg/day) Fecal N (mg/day) Urinary N (mg/day) N balance (mg/day)	14.50 ± 0.30 13.40 ± 1.60 8.60 ± 0.70 -8.60 ± 2.00

<sup>&</sup>lt;sup>a</sup> Values are expressed as mean ± S.E.M.

haemalun-eosin. The lengths of the villi and adjacent crypts were measured by means of a light microscope (Nikon 104, Charanton le Pont, France) equipped with a projective appa-

### Amino acid analysis

The amino acid determination of nitrogen sources was performed after an acid hydrolysis using 6N HCl at 110° C for 24 hr in an air-oven and under nitrogen at atmospheric pressure; then HCl was evaporated under reduced pressure at 40° C; tryptophan content was measured after alkaline hydrolysis using 4N NaOH at 110° C for 16 hr and under nitrogren atmosphere according to Hugli and Moore. 25 Amino acid analyses were then performed using a high-performance liquid chromatography (HPLC) system (Kontron SFM 25, Zurich, Switzerland) with a Shimadzu integration system. Derivatization procedure used 9-fluorenylmethylchloroformate (FMOC-Cl). All the subsequent steps were carried out according to Einarsson.<sup>26</sup>

### Statistical analysis

Results are given as mean ± SEM. Data were compared by analysis of variance (ANOVA); significance was determined by a multiple range Student's t test.

### Results

The effects of malnutrition are summarized in Table 3; the animals lost 29% of their body weight during 16

days of malnutrition. The mean total protein intake represents an amount of protein too low to meet the protein requirement of the adult rat (approximately 1.8–2.0 g/day). On the basis of nitrogen excretion, the animals were in a severe negative balance. It must be pointed out that urinary N was lower than fecal N, giving evidence of the malnutrition status of the rats.

Table 4 shows the nutritional parameters of rats after rehabilitation. The initial body weight differs slightly from the final body weight in the control group (Table 3) because five animals were killed at random before the restoration period; therefore, means were modified. Refeeding DTP diet produced a pronounced growth. This diet had the best food efficiency although no significant difference was observed between the three groups in terms of food intake; refeeding BP or AAM failed to reach such an efficiency. Nitrogen retention followed a similar pattern. Nitrogen digestibility was lower in AAM refed rats; nutritional performances of these animals were weaker than those of the other groups and particularly in comparison with the DTP fed group. None of the three diets caused diarrhea.

Intestinal alkaline phosphatase activities (AP) are given in Figure 1. After refeeding, activities increased independent of the diet utilized. Activities were identical between BP and DTP refeds, these last values being significantly weaker than in AAM ones. In the ileal segment, a decrease in mucosal protein content dependent on the malnutrition status was observed (Table 5); refeeding short peptides resulted in maximal mucosal protein contents. Ileal AN activities expressed per total segment or per milligram of protein (Table 5) were significantly lower after malnutrition. Refeeding permitted a repair of enzymic activities, either segmental or specific. Examination of segmental activities revealed that the maximal AN activity was confined to DTP refeds. No discernible differences were noted between BP and DTP refeds concerning the specific activities, however, a weaker difference was noticeable in AAM refeds.

**Table 4** Nutritional parameters after refeeding\*

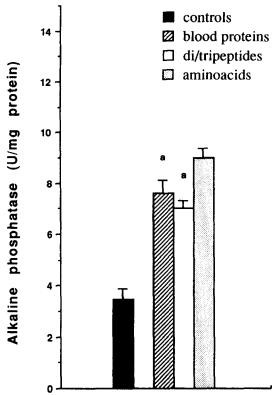
	Refeeding		
	ВР	DTP	AAM
Initial B.W. (g)	$80.6 \pm 2.5^{a}$	$80.6 \pm 2.5^{a}$	80.6 ± 2.0 <sup>a</sup>
Final B.W. (g)	$130.0 \pm 2.8$	$140.0 \pm 5.7$	107.4 ± 1.9
Growth (g/day)	$6.1 \pm 0.2$	$7.4 \pm 0.4$	$3.3 \pm 0.4$
Food intake (g)	$104.4 \pm 4.1^{a}$	$103.8 \pm 8.0^{a}$	$94.5 \pm 9.2^{a}$
Food efficiency†	$3.1 \pm 0.1$	$3.9 \pm 0.3$	$2.1 \pm 0.1$
N intake (mg/day)	$322.2 \pm 11.3^a$	315.2± 16.0 <sup>a</sup>	290.3 ± 8.8 <sup>a</sup>
Fecal N (mg/day)	$43.1 \pm 3.3^{ab}$	$40.1 \pm 3.3^{a}$	$46.0 \pm 2.5^{b}$
Urinary N (mg/day)	$125.6 \pm 4.8$	$112.7 \pm 6.1$	143.0 ± 5.1
N retention (%)±	$54.9 \pm 1.0$	$57.3 \pm 2.7$	$39.2 \pm 1.0$
CDU (%)§	$86.5 \pm 1.2^{a}$	$87.3 \pm 0.9^{a}$	82.3 ± 1.7

<sup>\*</sup> Values are expressed as mean  $\pm$  SEM; values in a same row with identical superscript letters are not significantly different (P < 0.05)

<sup>†</sup> Calculated as body weight gain/protein intake measured over the 8-day period.

<sup>‡</sup> Calculated as (N intake - Fecal N - Urinary N)/(N intake - Fecal N) measured during the last 3 days.

<sup>§</sup> Nitrogen apparent digestibility = (N intake - Fecal N)/N intake.



**Figure 1** Specific activities (units/mg protein) of alkaline phosphatase after malnutrition (controls, black bars) and after refeeding BP (hatched bars), DTP (clear bars), or AAM (stippled bars). Values represent means of five determinations per group  $\pm$  SEM; bars with the same superscript letter are not significantly different (P < 0.05).

Morphometric data are summarized in Figure 2; villus length and crypt depth were significantly lower after protein-calorie malnutrition, accounting for a severe nutrient deprivation. Refeeding DTP was associated with the greatest illus height; crypt depth being different among the three groups.

### Discussion

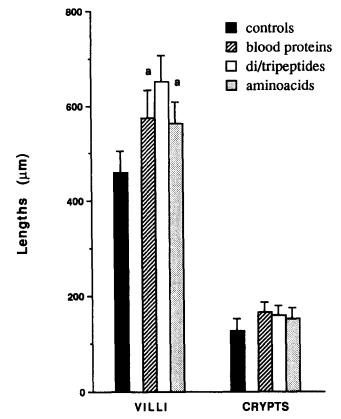
It seems likely that peptide-rich enzymic protein hydrolysates behave differently from equivalent amino acid mixtures when administered per os, in relation to either absorptive or postabsorptive events. 4-6.27 Few studies attempted to detect differences between them in terms of overall nutritional parameters, including food efficiency, nitrogen retention, and digestibility or in terms of intestinal behavior. So, the present experiments were designed to provide a basis for optimal utilization of either a dietary protein, its partial hydrolysate, or the corresponding constituent amino acids.

Data from Table 4 shows that when protein-calorie malnourished animals were refed at a standard nitrogen level in the diet (2.4% N), only DTP emerged from the three diets in terms of growth, food efficiency, and nitrogen retention. Moreover, rats receiving AAM diet had significantly greater nonfecal nitrogen losses, via increased urinary excretion of unchanged amino acids;

**Table 5** Effects of malnutrition and refeeding on ileal aminopeptidase activity and mucosal protein content<sup>a</sup>

	Mucosal protein (mg/segment)	U/segment	Specific activity (mU/mg protein)
Controls Refeds	18.2 ± 2.2	1.06 ± 0.1	55.0 ± 4.9
BP DTP AAM	$31.3 \pm 4.5^{a}$ $33.9 \pm 4.0$ $31.3 \pm 4.4^{a}$	$3.9 \pm 0.4$ $4.4 \pm 0.3$ $3.3 \pm 0.3$	$124.5 \pm 6.9^{a}$ $126.8 \pm 4.5^{a}$ $106.2 \pm 3.4$

<sup>&</sup>lt;sup>a</sup> Values with a same superscript letter in a column are not significantly different (P < 0.05); comparisons were made separately for each malnutrition schedule.



**Figure 2** Intestinal histomorphometric study after malnutrition and refeeding. Diets are indicated in Fig. 1. Three sections were examined for each specimen (ie, per rat); fifteen measurements were made per section. Bars with the same superscript letter are not significantly different (P < 0.05).

it is likely that the body of such animals cannot utilize all the dietary nitrogen for protein synthesis, suggesting an adaptive response to the moleuclar form of nitrogen supplied. In another respect, proteins are known to be the best substrate of intestinal proteolytic enzymes; replacing BP or DTP by AAM in the diet would be expected to modify the metabolism of endogenous nitrogen as well as the absorption of amino acids from the gut. The poor nitrogen digestibility of amino acid diet is not surprising; we have recently observed (unpublished data) that healthy rats fed 15% AAM diets ad libitum had a four-fold increase in intes-

tinal <sup>3</sup>H-thymidine incorporation compared to DTP or BP diets, providing a large enterocyte turnover and a subsequently increased rate for endogenous fecal nitrogen. If true in malnourished rats refed amino acid mixture, (high fecal nitrogen excretion), this might explain the slight coefficient of digestive utilization (CDU) observed. These nutritional data suggested that di- and tripeptides were utilized more efficiently than their equivalent amino acids mixture in rehabilitation of protein-calorie malnourished subjects.

We showed that nutritional rehabilitation of malnourished rats resulted in AP recoveries. The mechanisms of restoration seemed due to the nature of the nitrogen supply. The marked malnutrition status of rats before refeeding emphasized the burst in AP recovery observed in blood proteins and di/tripeptides refeds. Moreover, some authors<sup>29</sup> have shown that force-feeding rats with amino acids stimulated intestinal AP level; in relevance to this, restoration by feeding the amino acid mixture induced high levels of AP activity. Nevertheless, the role of AP in amino acid absorption remains unclear and the mechanism of the stimulation needs further elucidation.

The recovery in AN activities was also achieved after nutritional rehabilitation; nevertheless, these activities, both segmental and specific, varied markedly, depending on the diet administered. However, variations in segmental activities, which take into account mucosal changes and are closely related to the digestive capacity of the gut, were more remarkable than those of the specific activities. Our data suggested that there was a specific action of di- and tripeptides differing from that of proteins (BP) and total hydrolysis products (AAM) on AN activity. These adaptational enzymic differences not only appeared dependent on a specific substrate<sup>30,.31</sup> and different pancreatic secretions, but also on the nature and/or the molecular form of the dietary nitrogen. These findings were corroborated by the morphometric study. Similar results were recently reported<sup>32</sup> although experimental conditions such as protein source and malnutrition schedule differed from those used in this report. It remains for future investigation to elucidate the mechanisms involved in these changes. This work also indicates that the alterations of intestinal function and morphology as well as of nutritional parameters caused by malnutrition, can be rapidly reversed during a brief period of refeeding by di- and tripeptides. Increased demands for development of oligopeptides products for a therapeutic use justify such a research.

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### References

- 1 Grimble, G.H., Rees, R.G., Kehoane, P.P., Cartwright, T., Desreumaux, M., and Silk, D.B.A. (1987). Effect of peptide chain length on absorption of egg protein hydrolysates in the normal human jejunum. *Gastroenterology* 92, 136-142
- Steinhardt, H.J., Paleos, G.A., Brandl, M., Fekl, W.L., and Adibi, S.A. (1984). Efficacy of a synthetic dipeptide mixture as the source of amino acids for total parenteral nutrition in a subhuman primate (Baboon). Gastroenterology 86, 1562-1569
- 3 Solimano, G., Burgess, E.A., and Levin, B. (1967). Proteincalorie malnutrition: effect of diets on enzyme levels of jejunal mucosa of rats. *Br. J. Nutr.* 21, 55-68
- 4 Prosper, J., Murray, R.L., and Kern, F. (1968). Protein starvation and the small intestine. II. Disaccharidase activities. *Gastroenterology* **55**, 223–228
- 5 Adams, J.L., and Leichter, J. (1976). Effect of protein deficient diets with various amounts of carbohydrates on intestinal disaccharidases activities in the rat. J. Nutr. 103, 1716-1722
- 6 Lis, M.T., Matthews, D.M., and Crampton, R.F. (1972). Effects of dietary restriction and protein deprivation on intestinal absorption of protein digestion products in the rat. Br. J. Nutr. 28, 443-446
- 7 Raul, F., Noriega, R., Doffoel, M., Grenier, J.F., and Haffen, K. (1982). Modifications of brush border enzyme activities during starvation in the jejunum and ileum of adults rats. Enzyme 28, 328-335
- 8 Kim, Y.S., Mc Carthy, D.M., Lane, W., and Fong, W. (1973). Alterations in the level of peptide hydrolases and other enzymes in brush border and soluble fractions of rat small intestinal mucosa during starvation and refeeding. *Biochim. Biophys. Acta* 321, 262–273
- 9 Yamada, K., Goda, T., Bustamante, S., and Koldovsky, O. (1983). Different effects of starvation on activity of sucrase and lactase in rat jejunoileum. Am. J. Physiol. 244, G449-G455
- Syme, G. (1982). The effect of protein-deficient isoenergetic diets on the growth of rat jejunal mucosa. Br. J.Nutr. 48, 25-36
- 11 Syme, G., and Smith, M.W. (1982). Intestinal adaptation to protein deficiency. *Cell. Biol. Intern. Rep.* **6**, 573–578
- Hopper, A.F., Wannemacher, R.W., and Mc Govern, P.A. (1968). Cell population changes in the intestinal epithelium of the rat following starvation and protein-depletion. *Proc. Soc. Expl. Biol. Med.* 128, 695-698
- 13 Koga, A., and Kimura, S. (1979). Influence of restricted diet on the cell of the mouse small intestine. J. Nutr. Sci. Vitaminol. 25, 265-267
- 14 Lipscomb, H.L., and Sharp, J.G. (1982). Effects of reduced food intake on morphometry and cell production in the small intestine of the rat. Virchows Arch. (Cell Pathol) 41, 285-292
- 15 Misch, D.W. (1980). Intestinal microvilli: responses to feeding and fasting. Eur. J. Cell Biol. 21, 269-279
- Brunser, O. (1977). Effects of malnutrition on intestinal structure and function in children. Clinics in Gastroenterol., 6, 341-343
- Aldewachi, H.S., Wright, N.A., Appleton, D.R., and Watson, A.J. (1975). The effect of starvation and refeeding on cell population kinetics in the rat small bowel mucosa. J. Anat. 1, 105-121
- 18 Mac Manus, J.P.A., and Isselbacher, K.G. (1970). Effect of fasting versus feeding on the rat small intestine. Gastroenterology 59, 214-222
- 19 Gorostiza, E., Poullain, M.G., and Marche, C. (1985). Effect of fasting and refeeding on small intestinal adaptation in the rat. Gastroenterol. Clin. Biol. 9, 790-796
- 20 Kotler, D.P., Kral, J.G., and Björntorp, P. (1982). Refeeding after a fast in rats: effects on small intestinal enzymes. Am. J. Clin. Nutr. 36, 457-462
- 21 Bressollier, P., Petit, J.M., and Julien, R. (1988). Enzyme hydrolysis of plasma proteins in a CSTR ultrafiltration reactor: performances and modeling. *Biotechnol. Bioeng.* 31, 650-658
- 22 Bessey, O.A., Lowry, O.H., and Brock, M.J. (1946). Rapid colorimetric method for the determination of alkaline phospha-

- tase in five cubic millimeters of serum. J. Biol. Chem. 164, 321-329
- 23 Maroux, S., Louvard, D., and Baratti, J. (1973). The amino-peptidase from hog intestinal brush border. *Biochim. Biophys. Acta* 321, 282-295
- 24 Lowry, D.A., Rosebrough, N.F., Farr, A.L., and Randall, R.J. (1951). Protein measurement with the Folin phenol reagent. J. Biol. Chem. 193, 265-275
- 25 Hugli, T.E., and Moore, S. (1972). Determination of the tryptophan content of proteins by ion exchange chromatography of alkaline hydrolysates. J. Biol. Chem. 247, 2828–2832
- 26 Einarsson, S. (1986). Selective determination of secondary amino acids using precolumn derivatization with 9-fluorenylmethylchloroformate and reversed-phase high-performance liquid chromatography. J. Chromatog. 348, 213-220
- 27 Hegarty, J.E., Fairclough, P.D., Moriarty, K.J.J., Clark, M.L., Kelly, M.J., and Dawson, A.M. (1982). Comparison of plasma and intraluminal amino acid profiles in man after meals containing a protein hydrolysate and equivalent amino acid mixture. Gut 23, 670-674

- 28 Kimura, H., and Arai, S. (1988). Oligopeptide mixtures produced from soy protein by enzymatic modification and their nutritional qualities evaluated by feeding tests with normal and malnourished rats. J. Nutr. Sci. Vitaminol. 34, 375-386
- 29 Triantaphyllopoulos, E., and Tuba, J. (1959). Changes in intestinal and serum alkaline phosphatase levels during absorption of certain amino acids. Can. Biochem. Physiol. 37, 711-719
- Nicholson, J.A., Mc Carthy, D.M., and Kim, Y.S. (1974). The responses of rat intestinal brush border and cytosol peptide hydrolase activities to variation in dietary protein content. Dietary regulation of intestinal peptide hydrolases. J. Clin. Invest. 54, 890-898
- 31 Zambonino Infante, J.L., Rouanet, J.M., Caporiccio, B., and Besançon, P. (1989). Effects of dietary protein and carbohydrate level in the rat small intestine: enzymic, histological and electron microscopy studies. Nutr. Rep. Int. 40, 313-321
- electron microscopy studies. Nutr. Rep. Int. 40, 313-321
  32 Poullain, M.-G., Cezard, J.-P., Marche, C., Roger, L., Mendy, F., and Broyart, J.-P. (1989). Dietary whey proteins and their peptides or amino acids: effects on the jejunal mucosa of starved rats. Am. J. Clin. Nutr. 49, 71-76